Supplemental Document

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Deep-learning-assisted microscopy with ultraviolet surface excitation for rapid slide-free histological imaging: supplement

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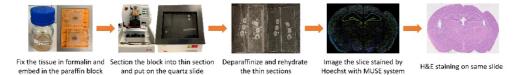


Fig. S1. Procedure for obtaining MUSE and H&E-stained images of thin mouse brain slices. A fresh mouse brain is formalin-fixed and paraffin-embedded in a block. Then, the paraffin block is sectioned and put on a quartz slide. After deparaffinization and rehydration, the tissue sample is stained with Hoechst and imaged by the MUSE system. To get the corresponding histological image, H&E staining is added on the same slice and viewed using a bright-field microscope.

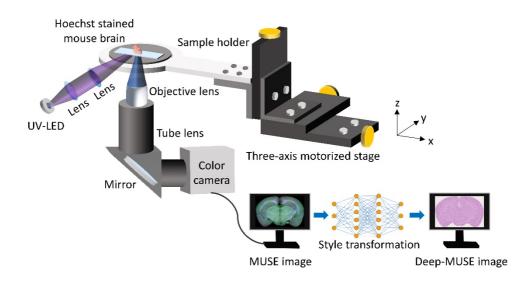
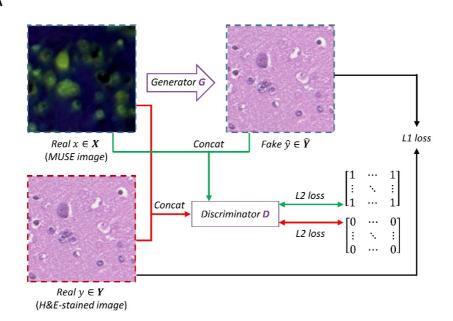


Fig. S2. Schematic of the MUSE imaging system integrated with a deep learning algorithm for rapid slide-free histological imaging. The optical configuration of the MUSE imaging system and style transformation from a MUSE image to a Deep-MUSE image that is comparable to a standard H&E-stained histological image.



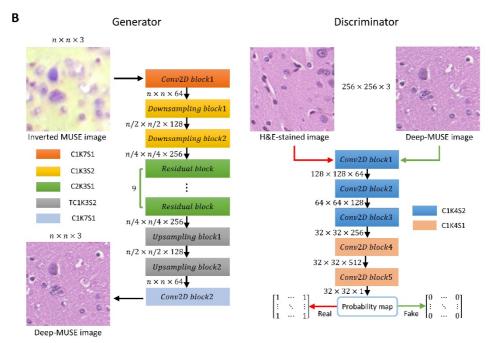


Fig. S3. The network architecture of the pix2pix and CycleGAN models. (A) The pix2pix model uses paired images for training. (B) The network architecture of the generator and the discriminator in both pix2pix and CycleGAN models. The tensor size of the input, intermediate, and output is represented by *Height* × *Width* × *Channel*, e.g., 256 × 256 × 3 denotes the RGB image with a pixel number of 256×256 . The color blocks have different structures and parameters, e.g., *C1K7S1* denotes the block has 1 convolution layer with 7×7 kernel size and stride 2, *TC1K3S2* denotes the block has 1 transposed convolution layer with 3×3 kernel size and stride 2.

Α

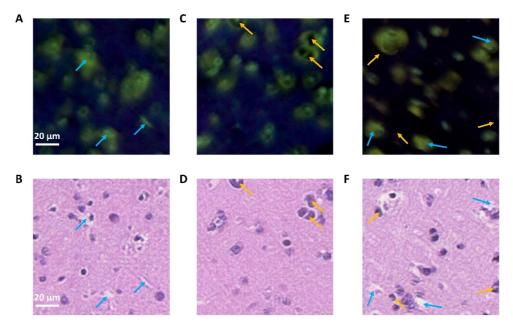


Fig. S4. Unsatisfactory style transformation of CycleGAN model training with original MUSE images without flipping contrast. (A, C, E) are three MUSE images of FFPE mouse brain slices. (B, D, F) are the Deep-MUSE images output by the CycleGAN model, corresponding to (A), (C), (E), respectively. The blue arrows show that the green cell nuclei in the MUSE images (A), (E) are converted into a white background in the Deep-MUSE images (B), (F). The orange arrows show that the black background in the MUSE images (C), (E) is converted into purple cell nuclei in the Deep-MUSE images (D), (F).

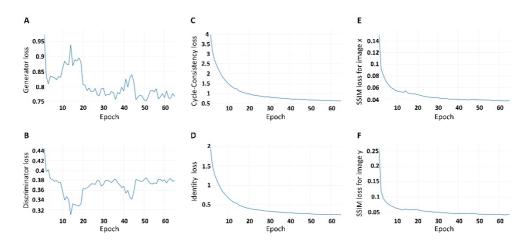


Fig. S5. Training loss curves of the CycleGAN model. (A) Generator loss curve of the generators G and F. (B) Discriminator loss curve of the discriminators D_X and D_Y . (C) Cycle consistency loss curve. (D) Identity loss curve. (E) SSIM loss curve of image domain X. (F) SSIM loss curve of image domain Y.

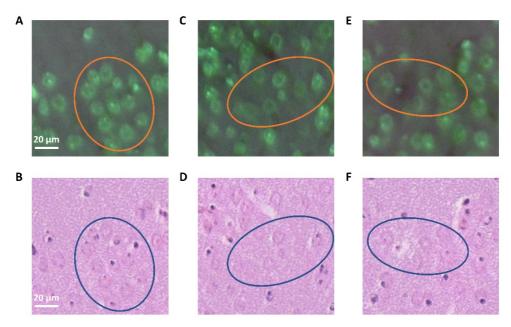


Fig. S6. Unsatisfactory style transformation by testing MUSE images of the thick tissue on a well-trained pix2pix model for FFPE thin slices. (A, C, E) are three MUSE images of a fixed and thick mouse brain. (B, D, F) are the Deep-MUSE images output by the pix2pix model, corresponding to (A), (C), (E), respectively. The blue circles in (B), (D), (F) show the missing cellular structures compared with the orange circles in (A), (C), (E).

Media S1. Closeup images of a column in the ROI of an FFPE mouse brain section acquired by MUSE, which are compared with their corresponding H&E-stained images using different methods. The 5X close-up scanning window with a pixel number of 500×500 (~221 × 221 μ m²). The images of different columns (from left to right) represent MUSE images, Deep-MUSE images by the pix2pix model, Deep-MUSE images by the CycleGAN model, and ground truth H&E-stained images.

Media S2. Closeup images of a column in the ROI of a fixed and thick mouse brain sample acquired by MUSE, which are compared with their corresponding Deep-MUSE images and the H&E-stained images of the adjacent layer. The 5X close-up scanning window with a pixel number of 500 \times 500 (~224 \times 224 μ m²). The images of different columns (from left to right) represent MUSE images, Deep-MUSE images by the CycleGAN model, and reference H&E-stained images of the adjacent layer.